## NOTES

## Ampullosporins B, C, D, $E_1$ , $E_2$ , $E_3$ and $E_4$ from *Sepedonium ampullosporum* HKI-0053: Structures and Biological Activities

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In a previous paper<sup>1)</sup> we reported the new peptaibol-type antibiotic ampullosporin A (1, Fig. 1), production by the fungal strain of *Sepedonium ampullosporum*, structure elucidation, and biological activities. Ampullosporin A (1) was discovered as an inducer of pigment formation by the fungal strain of *Phoma destructiva*. Moreover, strong neuroleptic activity was established for 1 in mice and rats as test animals<sup>1,2)</sup>.

In order to disclose structure-activity relationships amongst the naturally occurring homologues and isomers of ampullosporin A we investigated the metabolite spectrum of the producer strain *Sepedonium*  *ampullosporum* by HPLC and mass spectrometry (LC-MS, ESI-CID-MS/MS, ESI-CID-MS<sup>n</sup>).

These investigations unraveled the presence of minor components such as ampullosporins B (2), C (3), D (4) and E, amounting to  $1 \sim 5\%$  of the produced ampullosporin A (1). Component E was shown to be a mixture of four isomers (E<sub>1</sub> (5), E<sub>2</sub> (6), E<sub>3</sub> (7), E<sub>4</sub> (8); Fig. 1). Here we report the isolation, structure elucidation and biological activities of compounds  $2\sim 8$ .

The producer strain *Sepedonium ampullosporum* HKI-0053 was cultivated as was described previously<sup>1</sup>). Surface cultivation was carried out at 25°C in 1 liter Erlenmeyer flasks containing 100 ml of a medium composed of (g/liter): glycerol 30, glucose 10, peptone 5, NaCl 2, molecular sieve (5 nm, Merck) 1, agar 1; pH 7.0 (prior to sterilization).

After 14 days of cultivation as solid culture at 28°C the whole culture broth (40 liters) was extracted twice by 10 liters of ethyl acetate. The dried extract was evaporated and the residue was chromatographed on silica gel 60 (0.063~0.1 mm, CHCl<sub>3</sub>/MeOH, 9:1, v/v) and, subsequently Sephadex LH-20 (MeOH). The fractions containing ampullosporin A (1) and its congeners  $2\sim 8$  were detected by ESI-MS using m/z 1622 (1:  $[M+H]^+$ ) as a diagnostic tool. The mixture of crude ampullosporins (1.3 g) was separated subsequently by preparative HPLC

Fig. 1. Amino acid sequences of ampullosporins A (1), B (2), C (3), D (4),  $E_1$  (5),  $E_2$  (6),  $E_3$  (7) and  $E_4$  (8).

 $Ac-L-Trp^{1}-L-Ala^{2}-Aib^{3}-Aib^{4}-L-Leu^{5}-Aib^{6}-L-Gln^{7}-X-L-Gln^{11}-L-Leu^{12}-Y-L-Gln^{14}-L-Leuol^{15}-Leuol^{15}-L-Leuol^{15}-L-Leuol^{15}-L$ 

1	$X = Aib^8 - Aib^9 - Aib^{10}$	$Y = Aib^{13}$
2	$X = L-Ala^8 - Aib^9 - Aib^{10}$	$Y = Aib^{13}$
3	$X = Aib^{8}-L-Ala^{9}-Aib^{10}$	$Y = Aib^{13}$
4	$X = Aib^8 - Aib^9 - L - Ala^{10}$	$Y = Aib^{13}$
5	$X = L-Ala^8 - Aib^9 - Aib^{10}$	$Y = L - A la^{13}$
6	$X = Aib^{8}-L-Ala^{9}-L-Ala^{10}$	$Y = Aib^{13}$
7	$X = Aib^8 - Aib^9 - L - Ala^{10}$	$Y = L-Ala^{13}$
8	$X = L-Ala^8-L-Ala^9-Aib^{10}$	$Y = Aib^{13}$

Abbreviations: Ac-L-Trp: N-Acetyl-L-tryptophane, Aib:  $\alpha$ -Aminoisobutyric acid, L-Leuol: L-Leucinol, L-Ala: L-alanine.

on silica gel RP<sub>18</sub> (Nucleosil 100-A,  $40 \times 250$  mm, charged by 15 mg during each run with a gradient of 95% H<sub>2</sub>O to 95% acetonitrile, 25 minutes, 210 nm). Ampullosporins A~E were separable under these conditions due to differences in retention times (see Table 1). From 40 liters of the culture broth thus 0.95 g 1, 35 mg 2, 8 mg 3, 5 mg 4 and 3 mg of ampullosporins E (5, 6, 7, 8) were isolated as semicrystalline solid (Table 1).

Total hydrolysis of ampullosporins  $B \sim E$  (2~7) by 6 N hydrochloric acid, derivatization by Marfey's reagent and HPLC analysis<sup>3)</sup> suggested that they contained the same

amino acids and L-leucinol as  $1^{1}$ .

Structure elucidation of  $2 \sim 8$  was carried out by tandem mass spectrometry (triple quadrupole mass spectrometer Quattro, VG Biotech, Altrincham, England; LC-coupled ion-trap mass spectrometer LCQ, Finnigan, Bremen, Germany, both instruments equipped with electrospray ion source (ESI)) and NMR spectroscopy (<sup>1</sup>H, <sup>13</sup>C, DEPT 135, COSY, HSQC, HMBC, NOESY, TOCSY, Bruker AVANCE DRX 500).

The molecular weights of **2**, **3**, **4** and ampullosporin E (**5**, **6**, **7**, **8**) and chemical formulas were readily determined by

	2	3	4	5/6/7/8
Appearance	colorless solid	colorless solid	colorless solid	colorless solid
Melting Point (°C)	232-233 (decomposition)	157-159 (decomposition)	182-184 (decomposition)	222-224 (decomposition)
Molecular Weight	1607	1607	1607	1593
HRESI-MS ([M+Na] <sup>+</sup> )	1630.9284 (calcd. 1630.9272)	1630.9283 (calcd. 1630.9272)	1630.9281 (calcd. 1630.9272)	1616.9137 (calcd. 1616.9133)
Formula	C <sub>76</sub> H <sub>125</sub> N <sub>19</sub> O <sub>19</sub>	C <sub>76</sub> H <sub>125</sub> N <sub>19</sub> O <sub>19</sub>	C <sub>76</sub> H <sub>125</sub> N <sub>19</sub> O <sub>19</sub>	C <sub>75</sub> H <sub>123</sub> N <sub>19</sub> O <sub>19</sub>
$R_t$ (min) on HPLC (gradient H <sub>2</sub> O/acetoni- trile 1:99 to 99:1 16 min)	15.41	15.29	15.22	14.87

Table 1. Physico-chemical properties of ampullosporin B (2), C (3), D (4) and E (5, 6, 7, 8).

Table 2. Daughter ions observed with ampullosporins B (2), C (3), D (4) and  $E_1$  (5),  $E_2$  (6),  $E_3$  (7) and  $E_4$  (8) during CID-MS/MS and CID-MS<sup>n</sup>.

Ampullosporins	Fragments (m/z)											
	<b>B</b> <sub>1</sub>	B <sub>2</sub>	<b>B</b> <sub>3</sub>	<b>B</b> <sub>4</sub>	<b>B</b> 5	B <sub>6</sub>	<b>B</b> <sub>8</sub>	<b>B</b> 9	<b>B</b> <sub>10</sub>	B <sub>12</sub>	<b>B</b> <sub>13</sub>	$[M+H]^+$
1	229	300	385	470	583	668	881	966	1051	1292	1377	1622
2	229	300	385	470	583	668	867	952	1037	1278	1363	1608
3	229	300	385	470	583	668	881	952	1037	1278	1363	1608
4	229	300	385	470	583	668	881	966	1037	1278	1363	1608
5	229	300	385	470	583	668	867	952	1037	1278	1349	1594
6	229	300	385	470	583	668	881	952	1023	1264	1349	1594
7	229	300	385	470	583	668	881	966	1037	1278	1349	1594
8	229	300	385	470	583	668	867	938	1023	1264	1349	1594

For comparison the diagnostic daughter ions of ampullosporin A  $(1)^{i}$  are shown.

Residue	Proton	$\delta_{\rm H}$ (J in Hz)	δ <sub>c</sub>	Residue	Proton	δ <sub>H</sub> (J in Hz)	δ <sub>C</sub>
Ac	CH₃	1.85	22.63	Aib <sup>9</sup>	NH	8.00 (br, s)	
	CO		170.56		α		55.50
					$\beta_1$	1.34 (s)	22.39
Trp <sup>1</sup>	NH	8.32 (d, 7.5)			β2	1.39 (s)	26.61
	α	4.38 (ddd, 9.5, 7.5, 4.4)	54.81		ĊO		176.24
	βι	2.97 (dd, 15.0, 9.5)	27.14	. 10			
	$\beta_2$	3.11 (dd, 15.0, 4.4)		Ala <sup>10</sup>	NH	7.56 (br, d, 5.2)	
	1'	10.82 (d, 2.8)			α	3.90 (dq, 7.5, 5.2)	51.69
	2'	7.20 (d, 2.8)	123.69		β	1.42 (d, 7.5)	16.37
	3'		109.86		CO		174.94
	3a'		127.23				
	4'	7.53 (d, 7.9)	118.21	Gin''	NH	7.84 (br, d, 5.6)	
	5'	6.93 (t, 7.9)	118.12		α	3.96 (m)	55.17
	0	7.03 (t, 7.9)	120.88		$\beta_1$	1.96 (m)	26.17
	7	7.31 (a, 7.9)	111.33		$\beta_2$	2.04 (m)	
	/a CO		130.09		γ1	2.12 (m)	31.91
	0		175.15		γ2	2.34 (m)	
$\Delta la^2$	NH	8.38 (br. d. 5.3)			δ		173.41
1 114	a	4.02 (dg, 7.2, 5.3)	50.64		ε,	6.72 (br, s)	
	8	1.24 (d, 7.2)	15.96		ε2	7.18 (br, s)	
	CO	1.24 (0, 7.2)	174 29		CO		173.41
			174.29	12			
Aib <sup>3</sup>	NH	8.33 (br. s)		Leu <sup>12</sup>	NH	7.47 (d, 7.7)	
	α	0.00 (01, 0)	55 71		α	4.06 (m)	52.89
	<u>в</u> ,	1.35 (s)	22.30		βι	1.62 (m)	38.86
	B <sub>n</sub>	1.36 (s)	25 70		β <sub>2</sub>	1.64 (m)	
	CO		174 90		γ	1.69 (m)	24.13
			171.50		$\delta_1$	0.81 (d, 6.5)	21.07
Aib <sup>4</sup>	NH	8.01 (br. s)			$\delta_2$	0.84 (d, 6.7)	22.50
	a	0.01 (01, 5)	55 59		CO		172.91
	а В.	1 33 (s)	22.39				
	β.	1.37 (s)	26.79	Aib <sup>13</sup>	NH	7.48 (br, s)	
	C0	1.57 (5)	176.62		α		56.07
	00		170.02		βı	1.36 (s)	24.36
Leu⁵	NH	7 69 (d. 5 5)			β2	1.39 (s)	25.38
	α	3.88 (m)	54 60		CO		173.89
	а. В.	1.56 (m)	38.92	14			
	8- 8-	1 77 (m)	500,2	Gln'*	NH	7.16 (br, d, 7.8)	
	Ρ2 ν	1.71 (m)	24.51		α	3.97 (m)	53.33
	S	0.82 (4.6.6)	21.51		βι	1.80 (m)	27.14
	01	$0.82(\mathbf{u}, 0.0)$	21.45		$\beta_2$	1.99 (m)	· · · · · · · · · · · · · · · · · · ·
	02 CO	0.91 (0, 0.0)	22.30		γι	2.06 (m)	31.69
	co		174.08		γ2	2.19 (m)	
A :16	NH	7.83 (br. s)			δ		173.89
Alo		7.05 (01, 3)	55 78		ει	6.67 (br, s)	
	а. В.	133 (c)	23.73		ε2	7.06 (br, s)	
	PI B	1.55 (s)	25.25		CO		170.78
		1.45 (3)	175 76	T	NIT	7.02 (1.0.1)	
	co		175.70	Leuol	NH	7.02 (d, 8.1)	40.67
$Gln^7$	NH	7.37 (br. d. 5.4)			α	3.75 (m)	48.67
	<i>a</i>	3.80 (dt 7.0.54)	56.23		β <sub>1</sub>	1.31 (m)	39.77
	a. 8.	1.95 (m)	25.97		β <sub>2</sub>	1.34 (m)	(2.00
	рт В-	1 99 (m)	23.71		β <sub>1</sub> ΄	3.18 (dd, 10.4, 7.0)	63.89
	P2	2 11 (m)	31.03		$\beta_2$	3.30 (dd, 10.4, 5.0)	
	71 Va	2.11 (m) 2.20 (m)	51.05		Ŷ	1.63 (m)	23.93
	12 8	2.20 (11)	173 02		ð1	0.79 (d, 6.7)	21.77
	¢.	6.72 (br s)	175.02		δ <sub>2</sub>	0.83 (d, 6.7)	23.47
	51 52	7 19 (hr s)			· · · · · ·		
	CO		173.65				
A :1+8	NUT	8.02.4					
A10	NH ~	8.03 (br, s)					
	a	1 22 (a)	55.56				
	. p <sub>1</sub>	1.32 (8)	22.60				
	₽2 CO	1.45 (8)	25.80				
	CU		175.42				

Table 3. Assignments of the <sup>1</sup>H and <sup>13</sup>C NMR spectra of ampullosporin B (2) (DMSO- $d_6$ , 500 MHz,  $\delta$  in ppm, TMS as internal standard).

Abbreviations: s; singlet, d; doublet, t; triplet, q; quartet, m; multiplet, br; broad

HRESI-MS (high-resolution double focussing mass spectrometer Finnigan MAT 95XL).

As shown in Table 1 the ampullosporins B (2), C (3), and D (4) displayed the same molecular weight of 1607 Da suggesting that they are position isomers. Ampullosporin E was distinguishable by lower molecular weight (1593 Da, Table 1).

The sequence of amino acids in 2, 3 and 4 was determined unambiguously by ESI-CID-MS/MS (argon as collision gas) and FAB-MS (high-resolution sector-field instrument AMD-402, AMD Intectra, Bremen, Germany) furnishing diagnostic B-type fragments<sup>4)</sup> (Table 2). Collision-induced fragmentation (CID-MS/MS) of m/z1616.9 ([M+Na]<sup>+</sup>) of ampullosporin E suggested the occurrence of chromatographically inseparable isomers (5, 6, 7, 8; Table 2) due to the appearance of several series of fragments. As was shown for the related roseoferins<sup>5</sup>, the sequence of amino acids of peptide isomers such as  $E_1$  (5),  $E_2$  (6),  $E_3$  (7) and  $E_4$  (8) was assignable by ESI-MS<sup>n</sup> experiments (Finnigan LCQ, ion-trap analyzer, helium as collision gas). For instance, collision-induced dissociation of selected MS<sup>2</sup> daughter ions afforded several series of diagnostic MS<sup>3</sup> ions as a measure for the occurrence of position isomers (Table 2).

Additionally to the above experiments the structure of **2** as the major homologue of ampullosporin A (**1**) was confirmed by one- and two-dimensional NMR measurements. <sup>1</sup>H <sup>1</sup>H-COSY, HSQC, HMBC and NOESY correlations were particularly helpful for the assignment of the <sup>1</sup>H and <sup>13</sup>C signals and disclosure of the amino acid sequences (Fig. 1, Table 3).

Ampullosporins B, C, D,  $E_1$ ,  $E_2$ ,  $E_3$  and  $E_4$  (**2**~**8**) thus appear as new members of the peptaibol family of fungal peptides. Obviously, the structures of ampullosporins from *Sepedonium ampullosporum* show differences only in positions 8, 9, 10 and 13 whereby Aib is replaced by L-alanine.

The biological activities of **2**, **3** and **4** were compared with that of **1** using *Phoma destructiva* as test organism<sup>5)</sup>. In concentration of  $50 \,\mu\text{g/agar}$  well compounds **2** and **4** induced pigment formation in the same manner and concentration as ampullosporin A (**1**). However, compound **3** was needed in five-times higher concentration to afford a comparable effect.

The neuroleptic activity of  $2 \sim 4$  was tested in mice<sup>5)</sup>. After a dosage of  $1 \sim 10 \text{ mg } 2$  or 4 / kg body weight a decrease of body temperature was measured as a characteristic of neuroleptic drugs<sup>1)</sup>. The effect was comparable to that of ampullosporin A (1). In contrast, compound 3 caused a comparably much lower decrease of body temperature suggesting that sequence and quality of amino acids in positions 8, 9, 10 of the ampullosporins possess a crucial importance for biological activity as morphogenic and neuroleptic drugs.

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